



Continuous-cover forestry  
maintains soil fungal communities  
in Norway spruce dominated boreal forest

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# Decreasing Forests, Increasing Demands

Proportion and distribution of global forest area by climatic domain, 2020

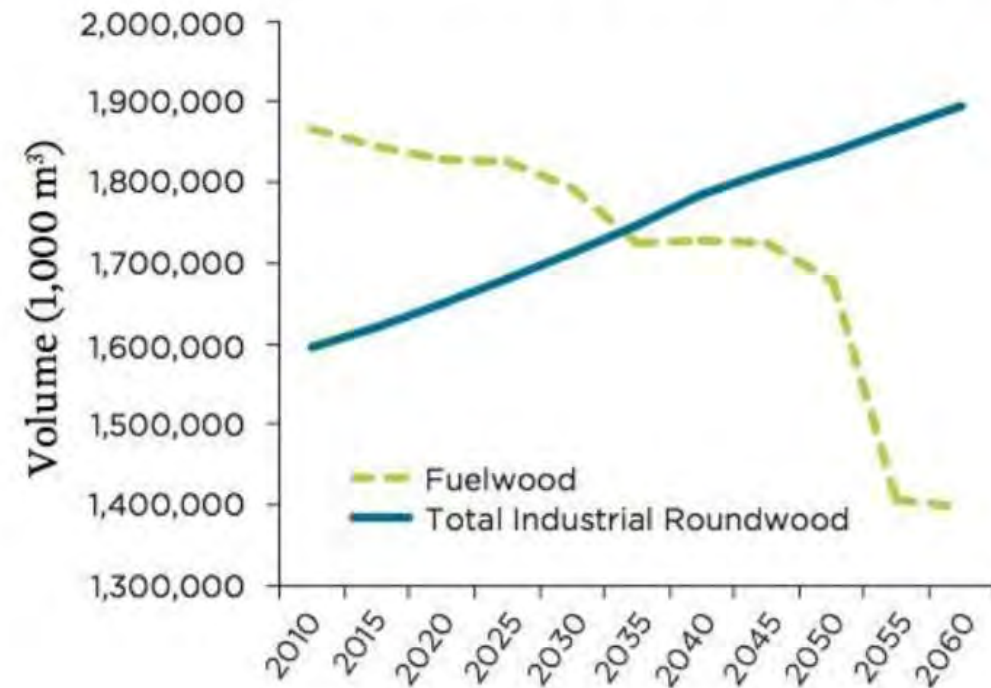


**The world's forest area has declined by 4.7M ha (> Denmark) a year since 2010 (FAO, 2020)**



Size of Denmark

Figure 1.16 Projected global consumption of wood, 2010–2060



Source: Pipa Elias and Doug Boucher, *Planting for the Future: How Demand for Wood Products Could Be Friendly to Tropical Forests* (Union of Concern Scientists, 2014).

# Why Boreal forests are important?



- ☐ Boreal forest?
  - Coniferous forests (pines, spruces, larches)
  - High-latitude environment
  - Freezing temperatures: 6-8months

- ☐ 11.5% of total land area.
- ☐ 1/3 global forest area
- ☐ 1/3 Carbon Stock
- ☐ Contains the most freshwater
- ☐ High ECM fungal diversity

Nilsson et al. 2019  
Pew environment group  
Dunn et al., 2007

- ☐ 1/3 timber, 1/4 paper in global trade
- ☐ Intensively managed
- ☐ N limited

Gauthier<sup>3</sup> et al. 2015

# Even-aged Forestry



**=Clear-fell forestry (CF)**

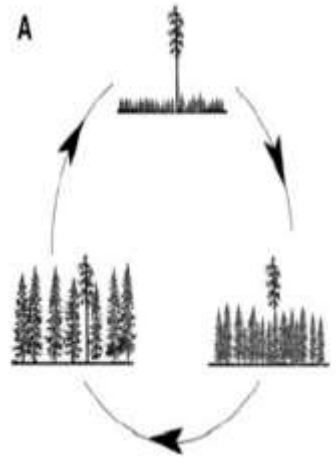
Harvests all trees in a given area (50~60 years)



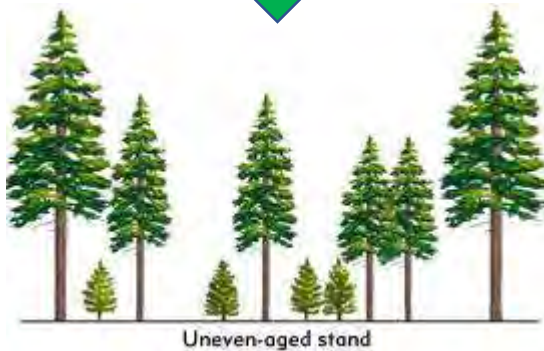
- ☐ Timber Production
- ☐ Artificial replantation



- ☐ Biodiversity, especially EMF
- ☐ Ecosystem function & services
- ☐ Less resilient
- ☐ Carbon storage



# Uneven-aged Forestry



**=Continuous-cover forestry (CCF)**

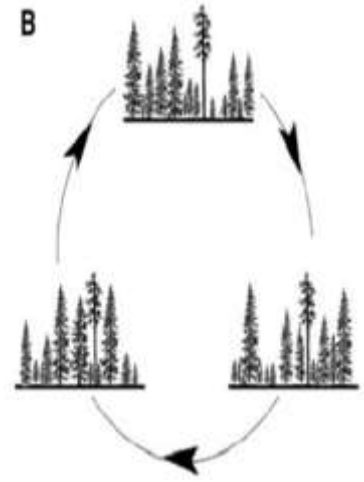
Harvests 30-40%, every 15-20 years



- ☐ Cost-efficient
- ☐ Diversity of plant & invertebrate
- ☐ Bilberry, mushroom production
- ☐ Resistance against wind



- ☐ Timber production
- ☐ Harder to manage



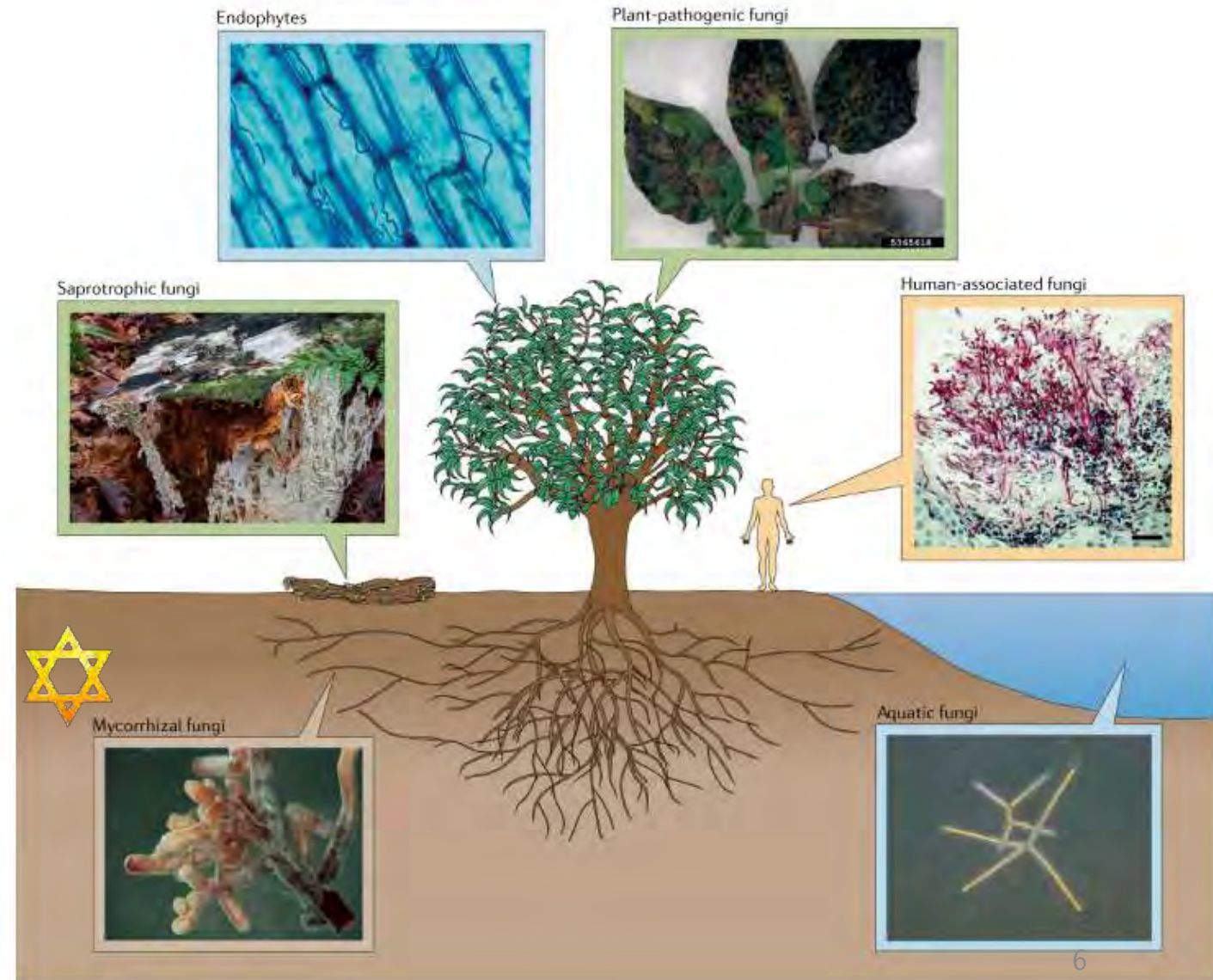
**We don't know about fungi!**

# Fungi

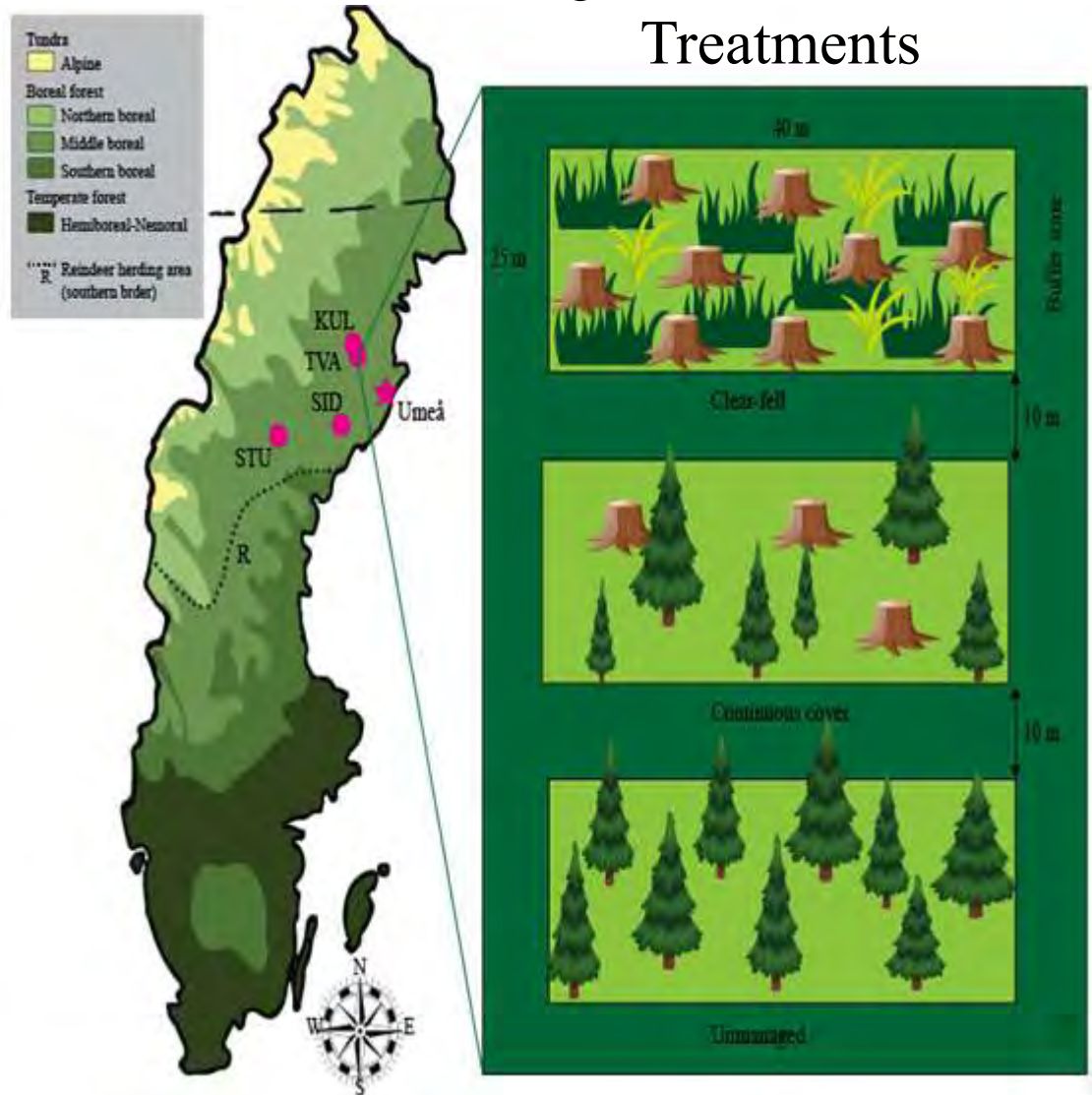
R.Henrik Nilsson et al. 2019

- **Saprotrophic fungi**  
mediate the decomposition of organic matter
- **Mycorrhizal fungi**  
associate with roots
- **Plant-pathogenic fungi**  
decompose living leaves
- **Endophytes**  
live inside plants and rarely visible to the naked eye  
Poorly understood nutritional strategies and taxonomic affiliations.

**Mycorrhizal fungi play a key role to provide nutrients to plants in boreal forest**



# Study Sites



❑ 4 sites established in 2012

❑ 3 treatments (Clear-fell, Continuous-cover, Unmanaged)

❑ Similar stands

Stand structure

Species composition

Age of forest

Norway spruce dominated

Field specificity

-Clear-fell condition: 100% cutting, replanted species, site preparation, stump retention

-Harvesting year:

2014-2015 for CF (100%) and CCF (30%) / UF (never logged).

Kulbäcksliden

CF VS CCF

Material&methods

Tvärålund

Sidensjö

Stugun

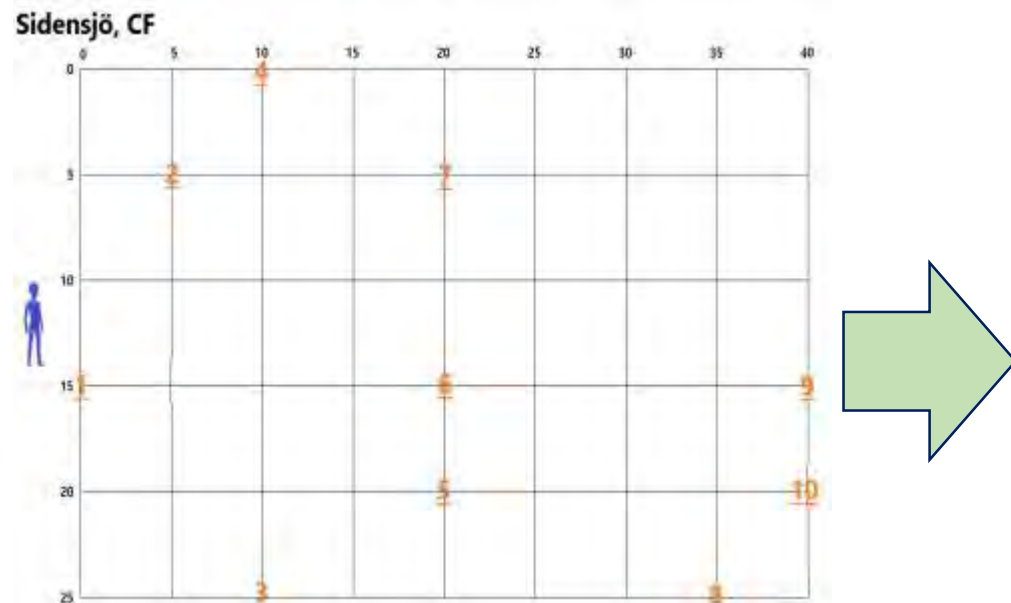


# Soil Sampling

4 sites \* 3 treatments \* 10 replicates = 120 soil samples

Soils for DNA Sequencing

Soils for Soil Chemical Analyses (pH, C, N, C/N)



# Sampling: Soil Chemicals



**pH meter**

the 5 ml of 0.01M CaCl<sub>2</sub>  
solution + 10s vortex



**Furnace for soil organic matter**

~550C for 6h at 550C for 6h  
~70C storing



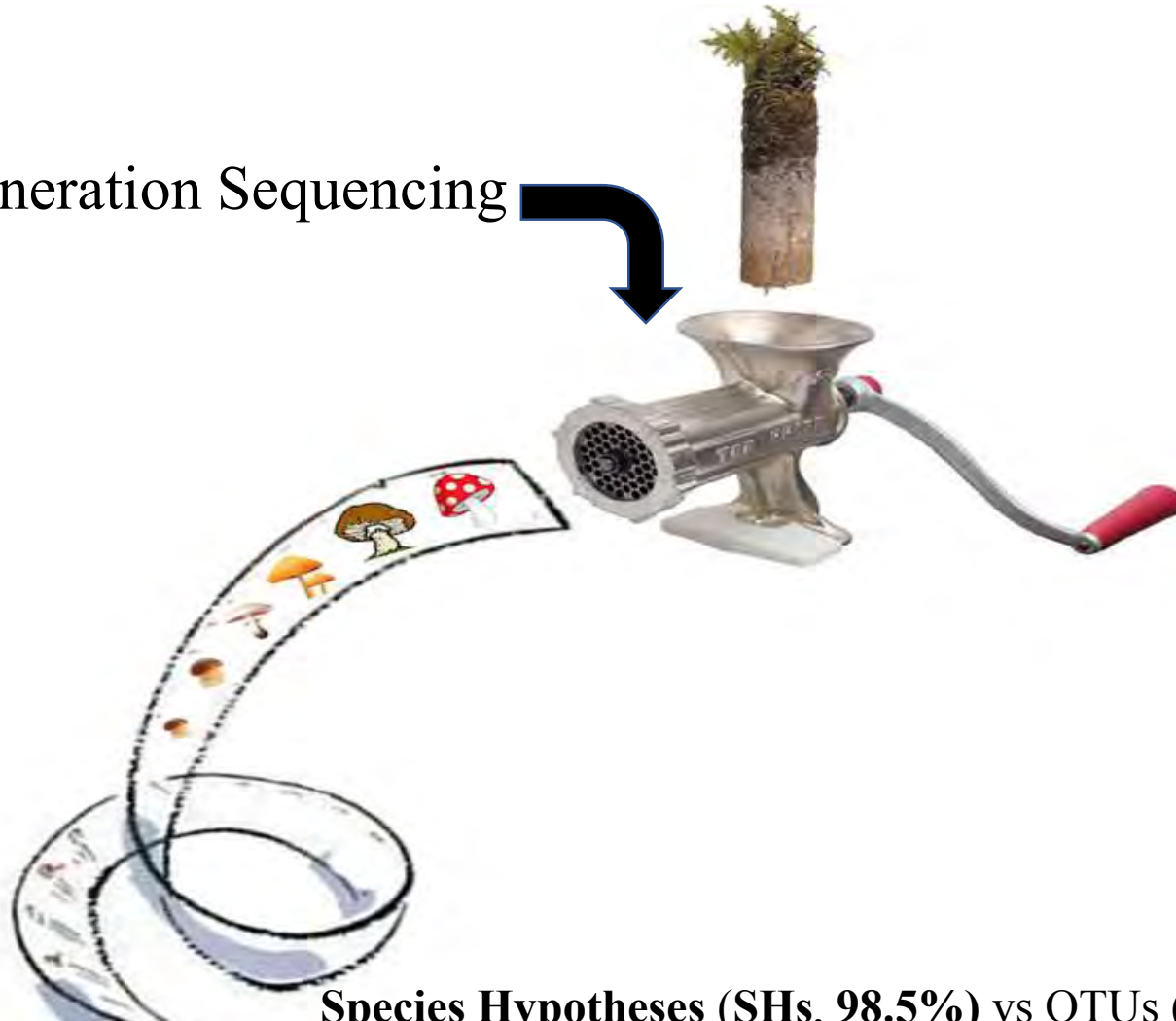
**Elemental Analyzer - Isotope Ratio Mass Spectrometer (EA-IRMS) for soil C, N and C/N**

Pre-treatment – freeze-dried & homogenization

# Sampling: Soil fungi

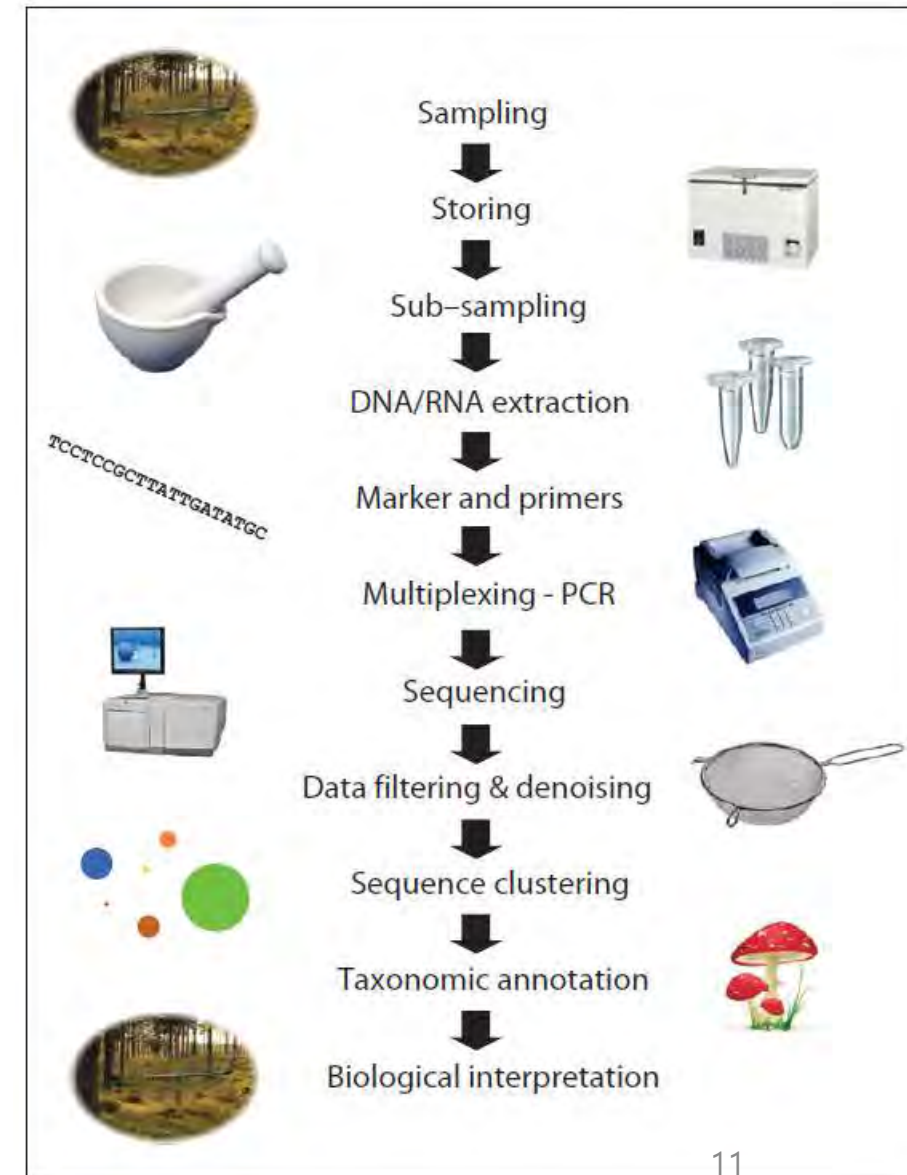
Imported by Björn Lindahl

Next Generation Sequencing

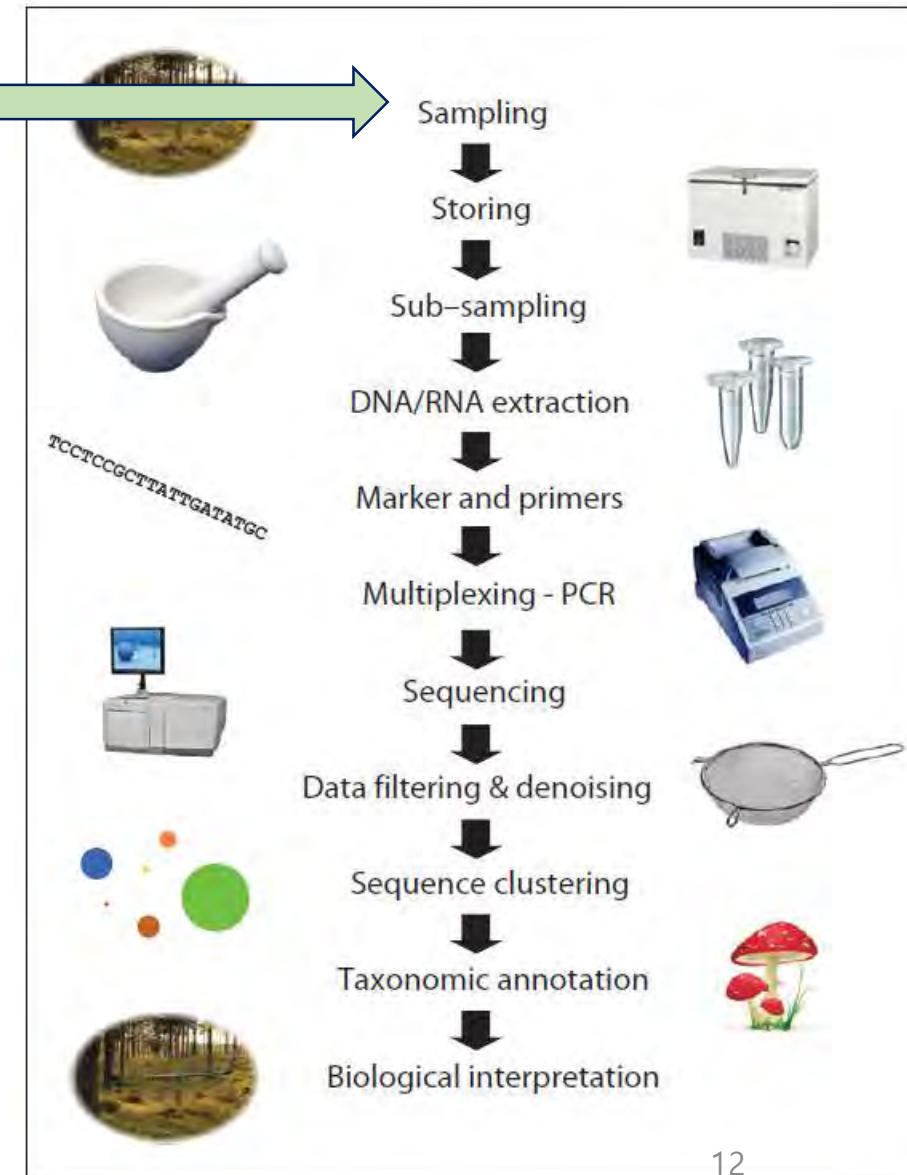


**Species Hypotheses (SHs, 98.5%)** vs OTUs (97%)  
: any species-level group of individuals that share a given set of observed characters

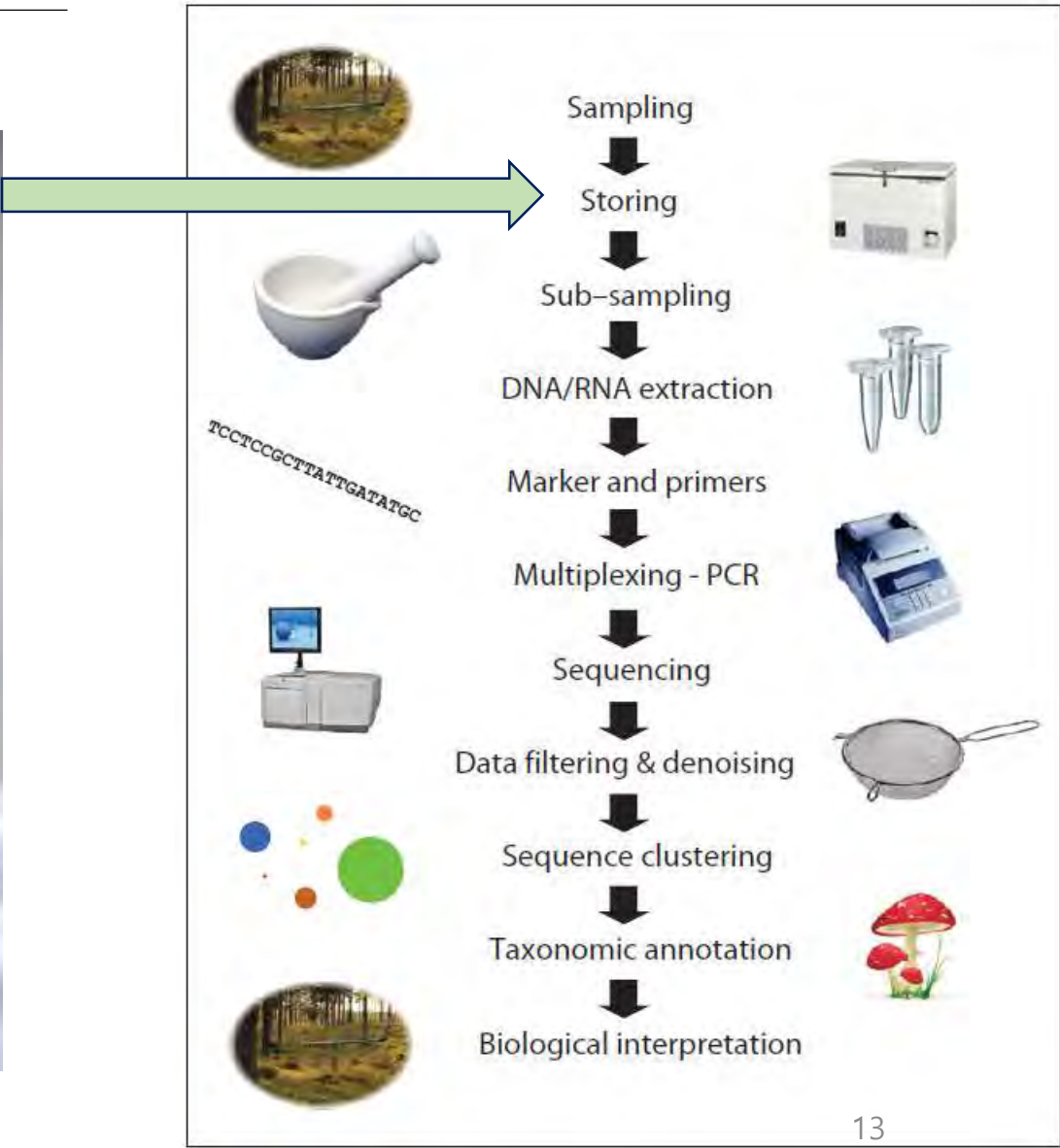
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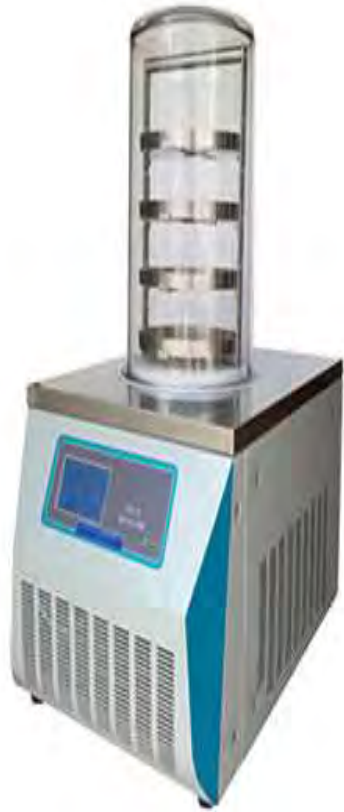
# Sampling: Soil fungi



# Sampling: Soil fungi



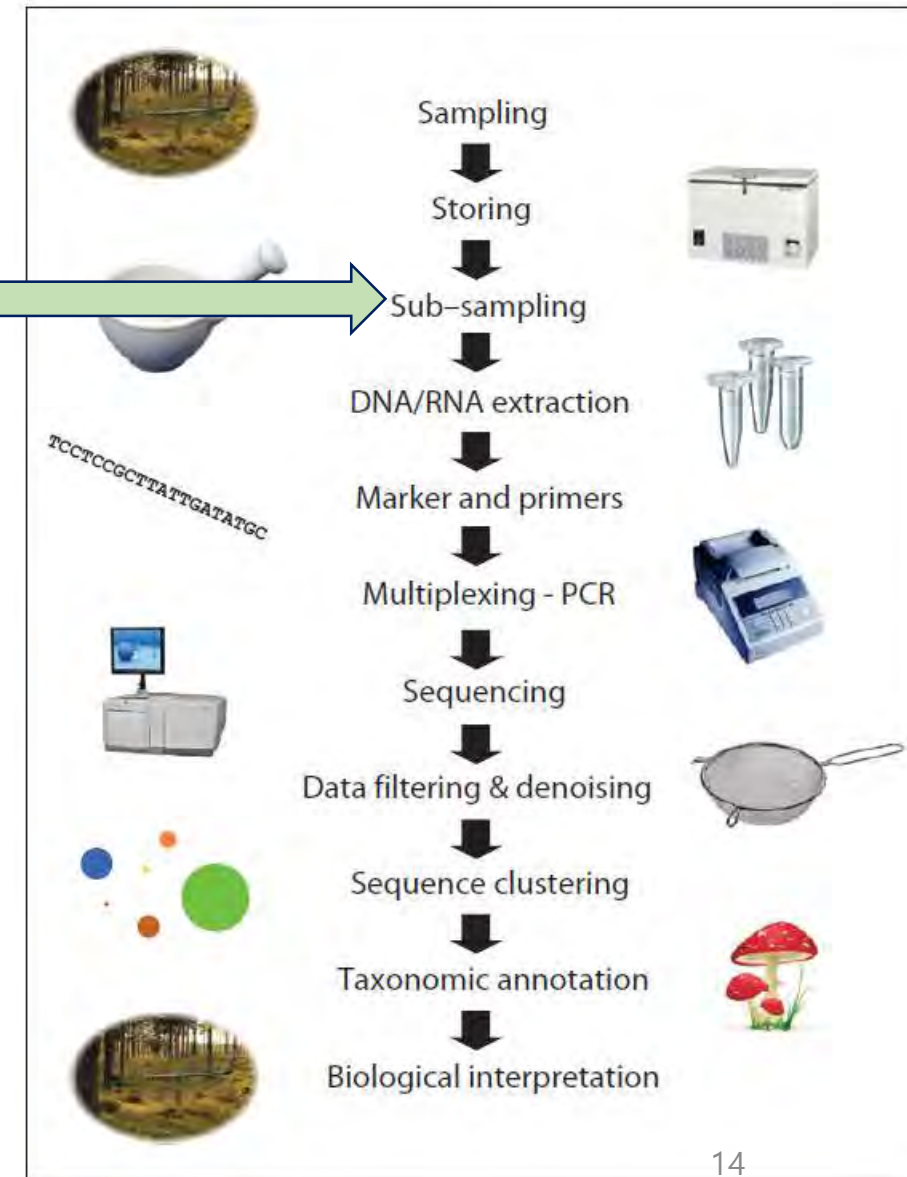
# Sampling: Soil fungi



Freeze-drier



Roller mill



# Sampling: Soil fungi

## Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for *Fungi*

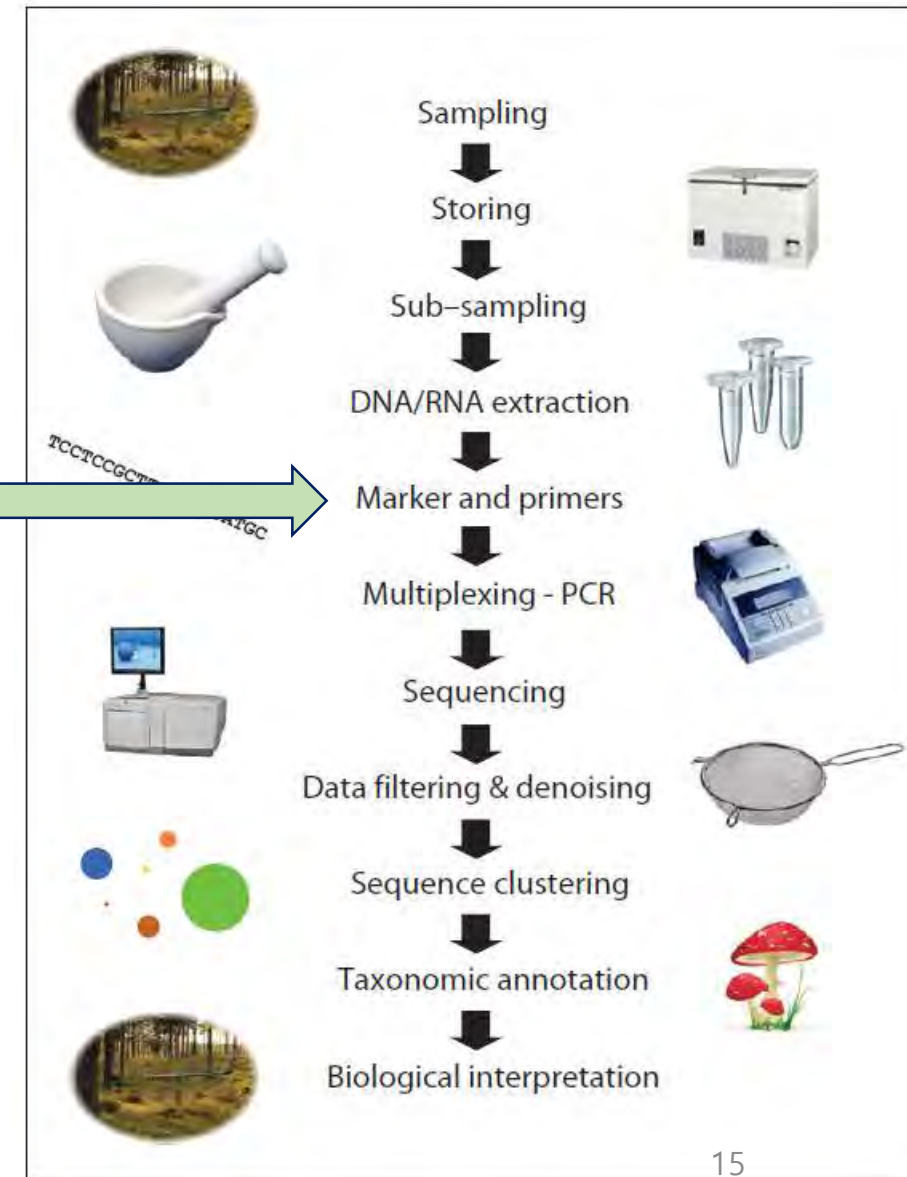
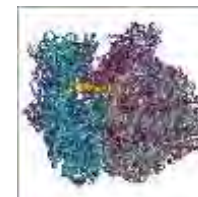
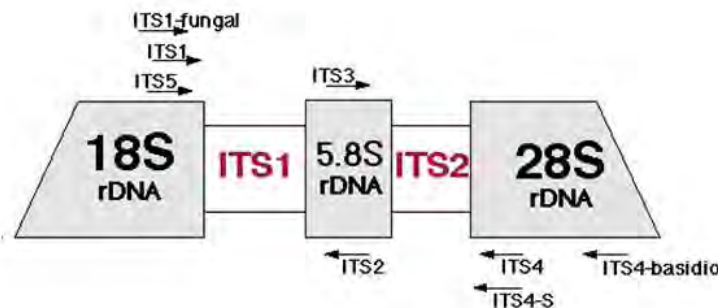
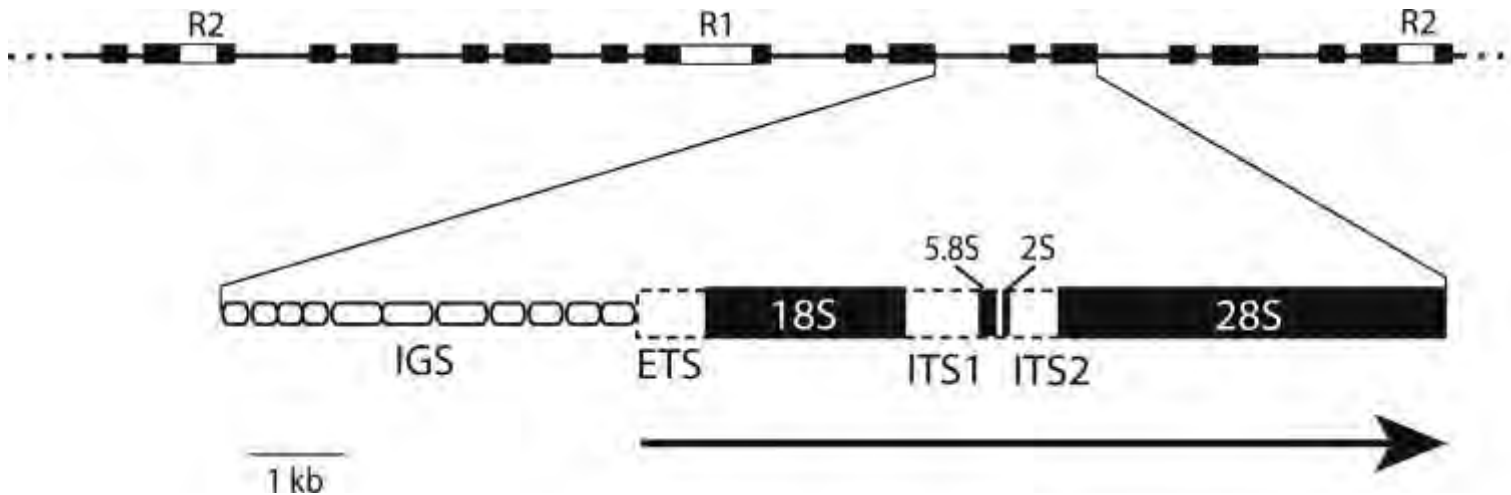
Conrad L. Schoch<sup>a,1</sup>, Keith A. Seifert<sup>b,1</sup>, Sabine Huhndorf<sup>c</sup>, Vincent Robert<sup>d</sup>, John L. Spouge<sup>a</sup>, C. André Levesque<sup>b</sup>, Wen Chen<sup>b</sup>, and Fungal Barcoding Consortium<sup>a,2</sup>

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Edited\* by Daniel H. Janzen, University of Pennsylvania, Philadelphia, PA, and approved February 24, 2012 (received for review October 18, 2011)

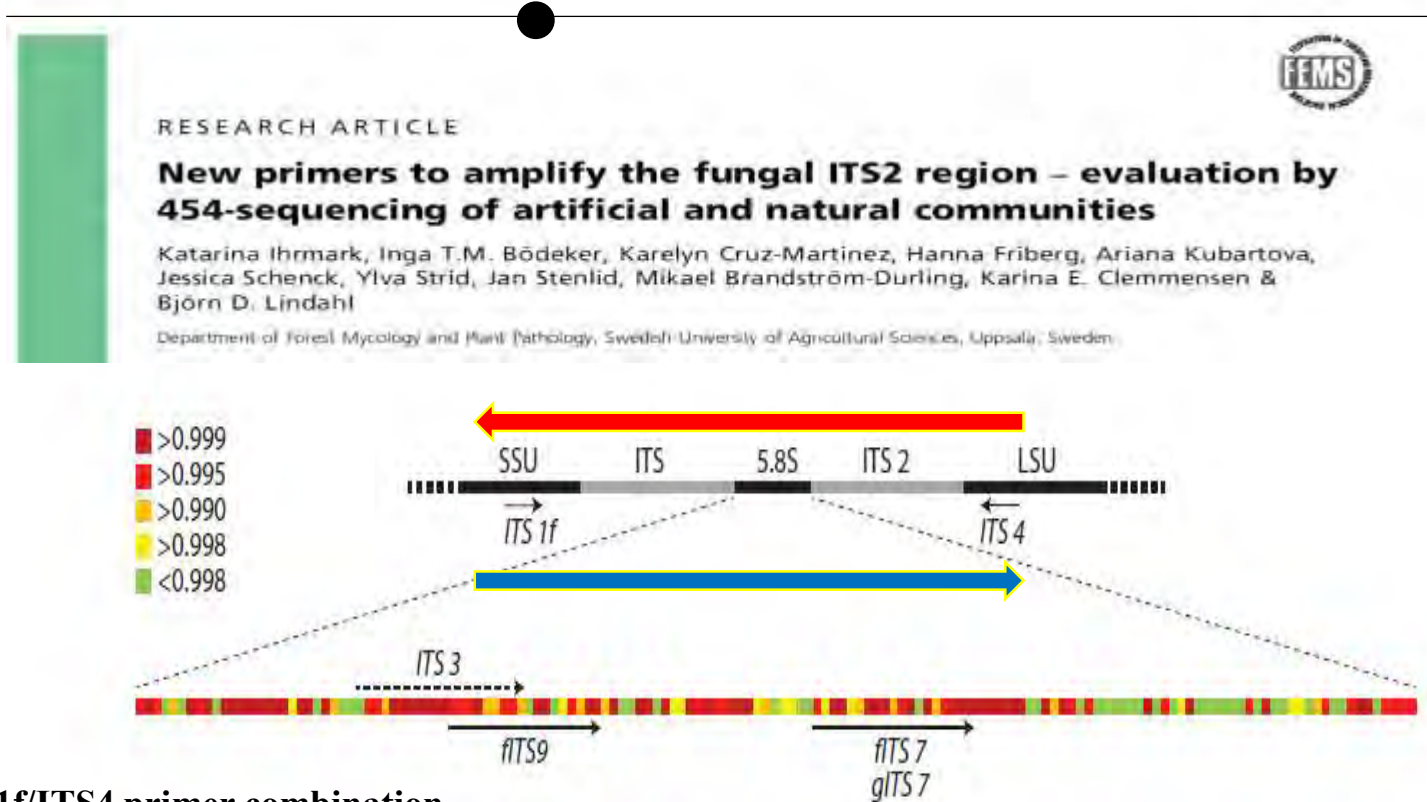


PNAS



- Routinely amplified
- Easy to detect
- A high degree of variation

# Sampling: Soil fungi



## ITS1f/ITS4 primer combination

+more genetic info

-strong bias against species with longer amplicons

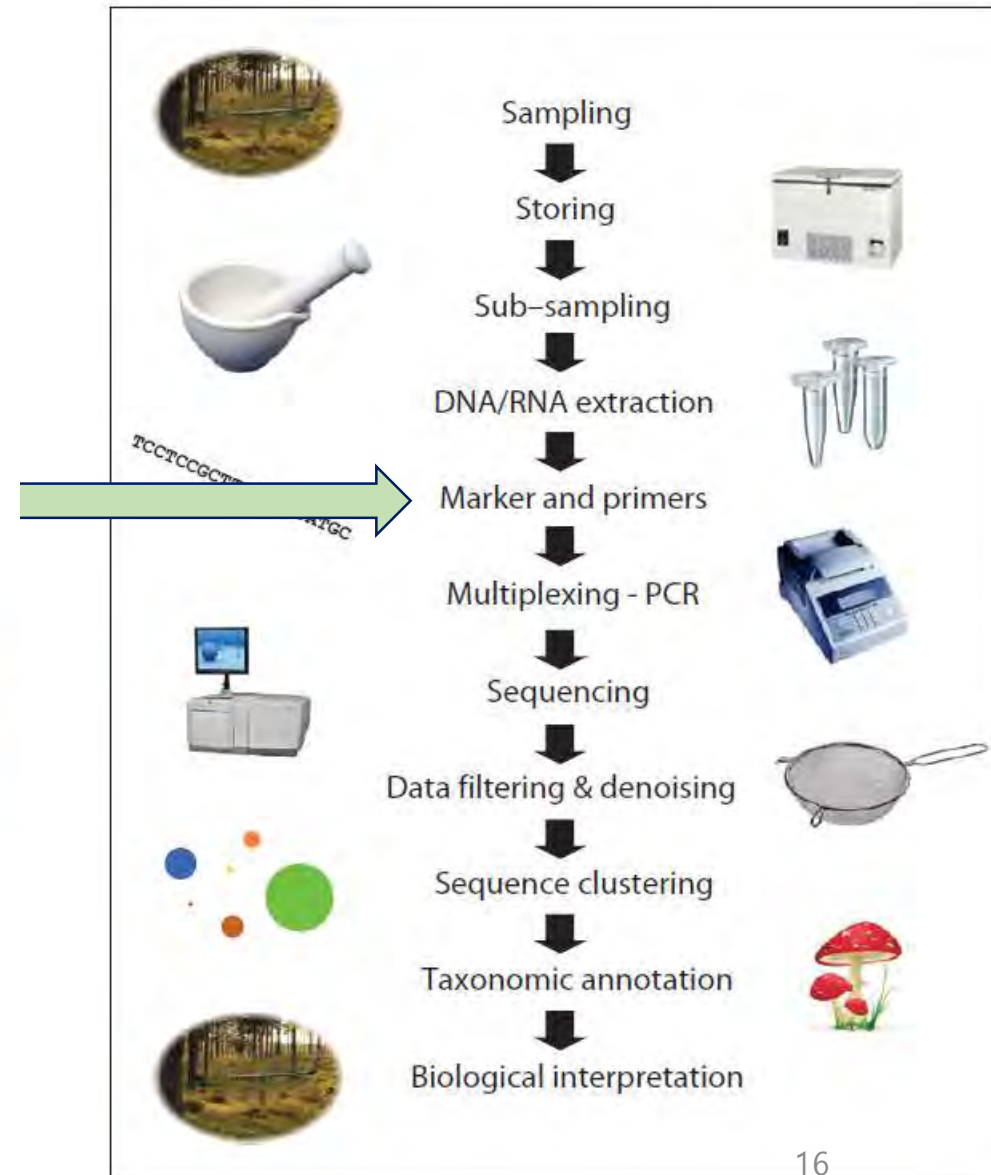
## New primer combination (gITS7 + ITS4)

+shorter amplicons and better preserve the quantitative composition of the template

+yielded more diverse amplicon communities

+higher PCR efficiencies

-less genetic info



# Sampling: Soil fungi

Illumina  
(MiSeq)

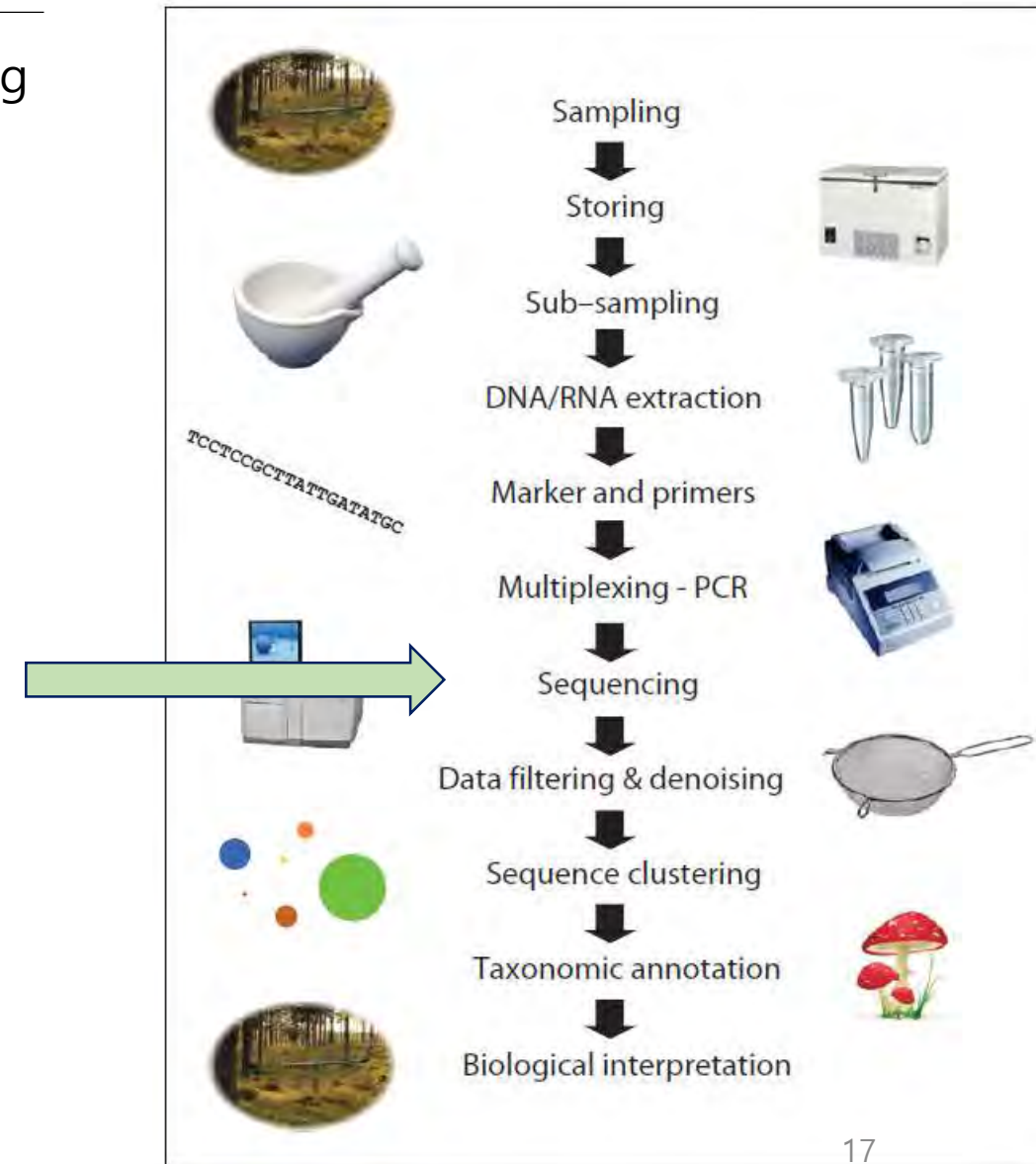


Pacific Biosciences SMRT sequencing  
(SEQUEL)

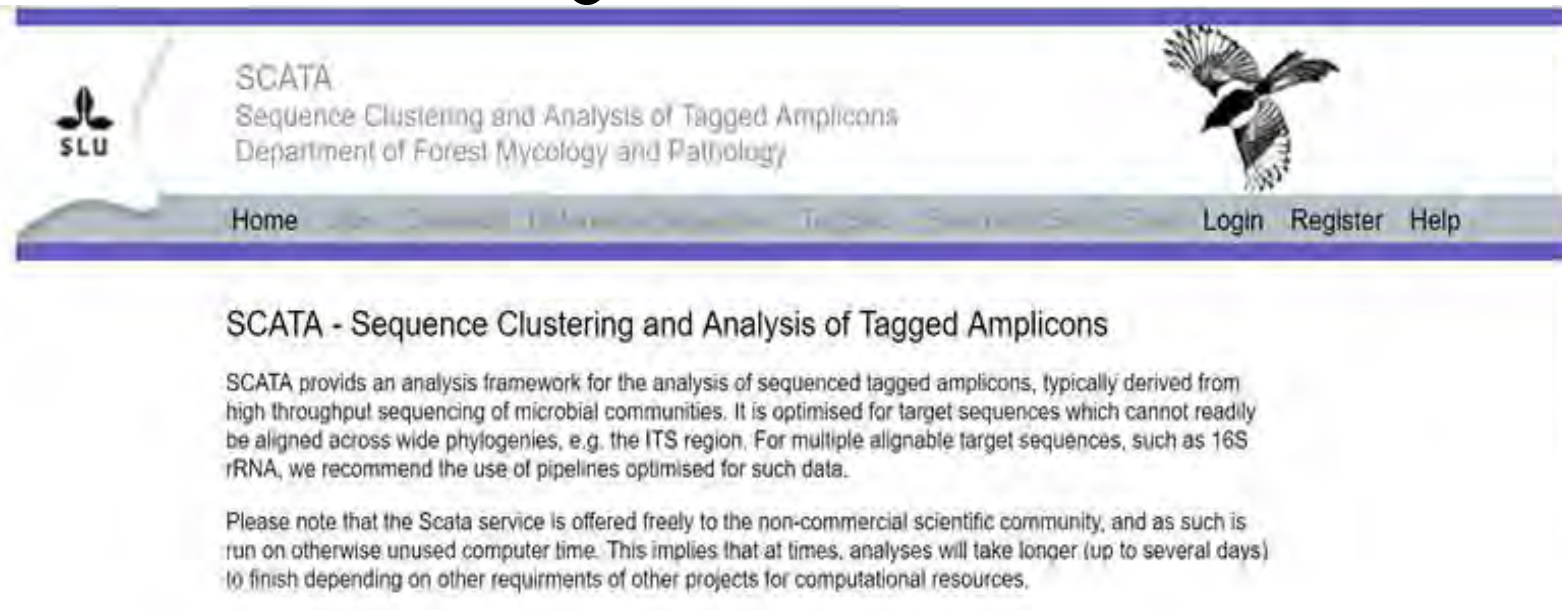


- 25 million sequencing reads
- 2 × 300 bp read lengths

- 500 000 sequencing reads
- 20 000 bp read lengths



# Sampling: Soil fungi



SLU

SCATA  
Sequence Clustering and Analysis of Tagged Amplicons  
Department of Forest Mycology and Pathology

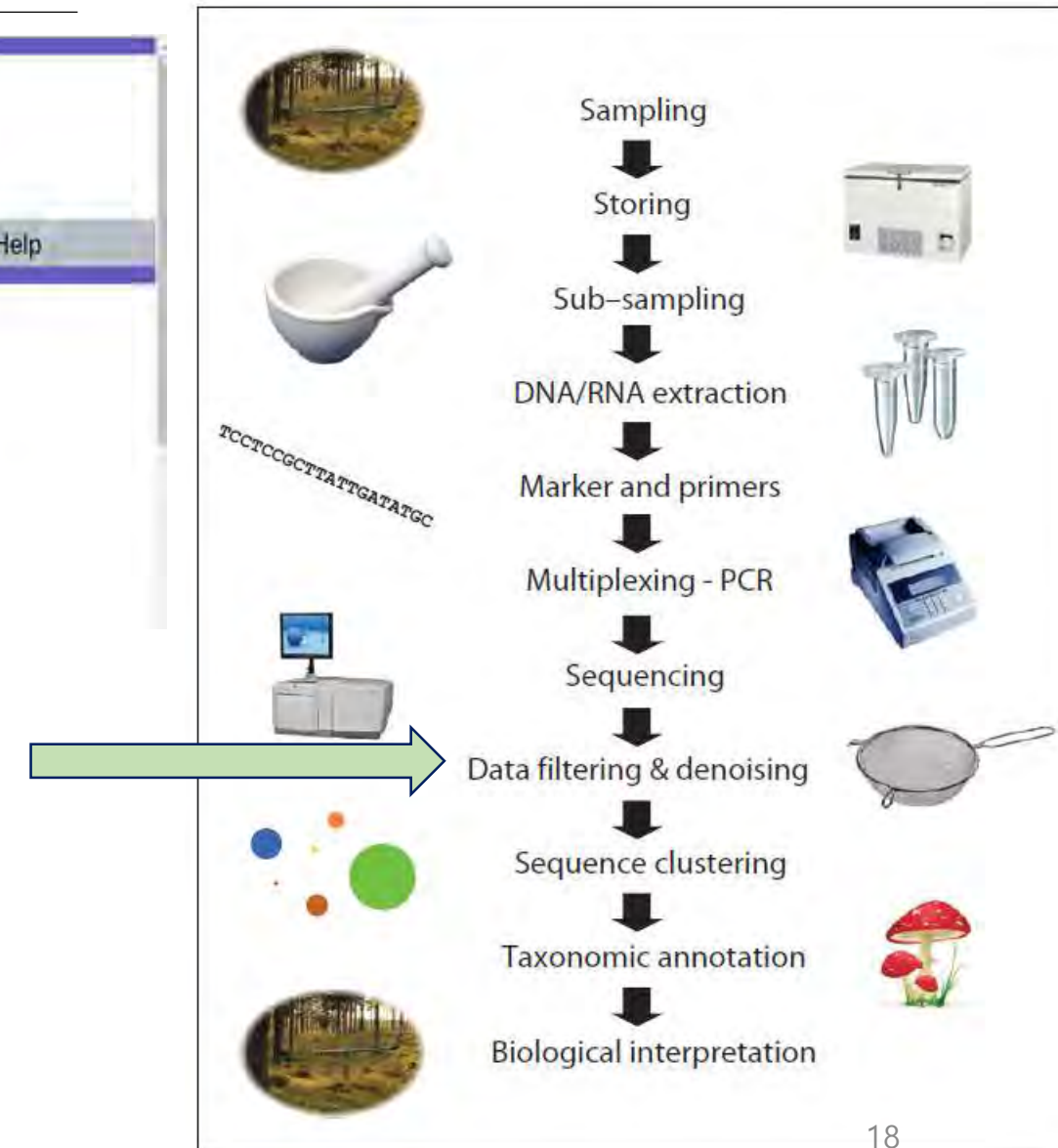
Home Login Register Help

## SCATA - Sequence Clustering and Analysis of Tagged Amplicons

SCATA provides an analysis framework for the analysis of sequenced tagged amplicons, typically derived from high throughput sequencing of microbial communities. It is optimised for target sequences which cannot readily be aligned across wide phylogenies, e.g. the ITS region. For multiple alignable target sequences, such as 16S rRNA, we recommend the use of pipelines optimised for such data.

Please note that the Scata service is offered freely to the non-commercial scientific community, and as such is run on otherwise unused computer time. This implies that at times, analyses will take longer (up to several days) to finish depending on other requirements of other projects for computational resources.

- Quality control
- Remove data with mean quality score below 20 or bases of equality lower than 3
- Screen gITS7 primer and identity tags
- Comparing sequences for similarity from USEARCH (match length – min. 85%)



# Sampling: Fruiting body

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D5 (CF, SID)



T17 (CCF, TVA)



S22 (UF, STU)

# Sampling: Fruiting body



## -Mushroom / Moss Identification

- >Visual & Microscopic Inspection
- >15 Mushroom illustrated books
- >5 Fungal DB
- >Herbarium of UmU

## -Dry mushrooms to measure biomass (50C, 72h)

= (6 events \* ID 4h) + Web-ID + Drying  
+ Cleaning + Measuring + Typing...

# Statistics

**H1. Soil fungal communities in CCF may be more similar to UF than CF**

**H2. Variation in soil fungal communities is related to soil chemical properties**

Software / Analysis	Univariate	Multivariate	Visualization
R studio	<ul style="list-style-type: none"> <li>- Mixed effects model</li> <li>- Tukey's HSD pair-wise</li> <li>- Correlation</li> </ul>		<ul style="list-style-type: none"> <li>-Anova table</li> <li>-Scatter plots</li> </ul>
PRIMER		<ul style="list-style-type: none"> <li>-PERMANAOVA</li> <li>-SIMPER</li> <li>-Diversity indices</li> <li>-DistLM</li> </ul>	<ul style="list-style-type: none"> <li>-SIMPER table</li> <li>-Canonical analyses of principal coordinates</li> <li>-DistLM table</li> </ul>

# Results of Soil fungi

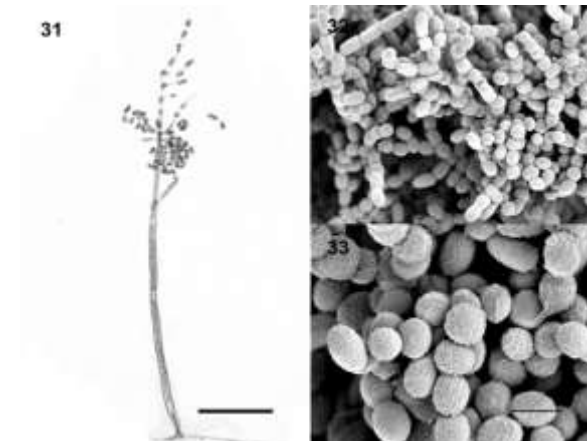
## Sequencing output

-549,011 reads -> 355,867 reads  
(65% after QC)

-1,016 reads per sample (SD: 142)

## -2157 Species Hypotheses

- 5 most common species (9% of total SHs)
- *Penicillium austroafricanum*
- *Hyaloscyphaceae* (Family)
- *Oidiodendron pilicola*
- *Luellia* (Genus)
- *Solicoccozyma terricola*



# Results of Fruiting body

- 1825 mushrooms (Identified 93.8%, 325g)
- 120 total species (excluding unidentified fungi)
- 43 Mycorrhizal fungi (145g)
- 3 most common species



Microphale perforans

UF CCF



Mycena epipterygia

UF CCF

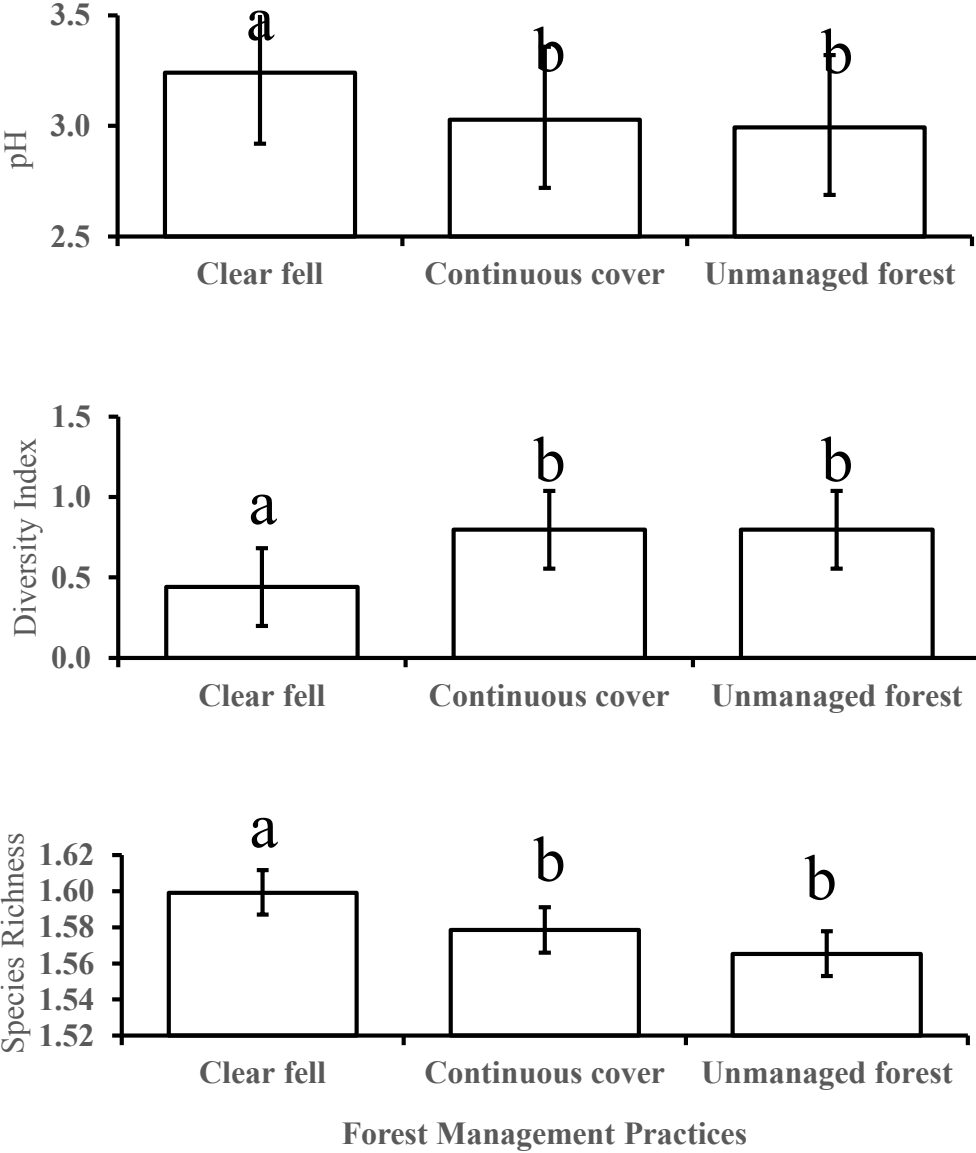


Cortinarius sp.

CF

# Mixed Effects Models

## 1) Impacts of forest management

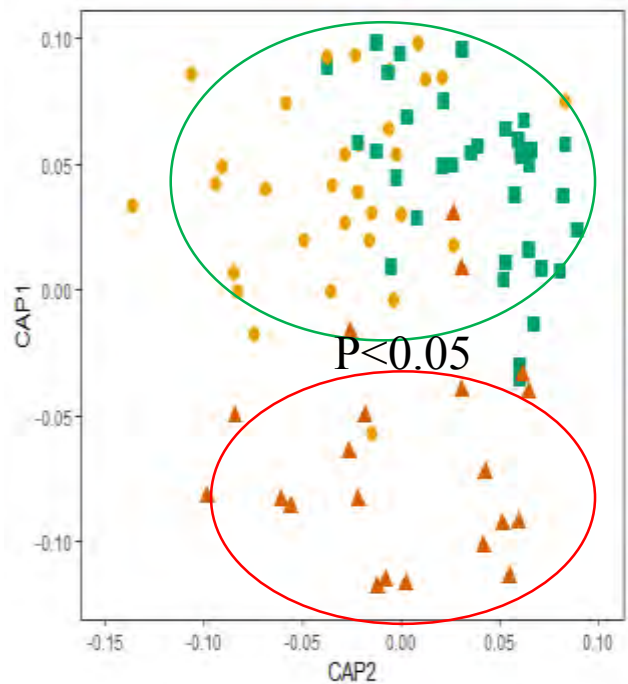


Dataset	Variable	F <sub>(df)</sub>	P-value	Clear fell	Continuous cover	Unmanaged
Soil chemistry	pH <sup>***</sup>	12.9 <sub>1,114</sub>	<0.001	3.2 <sup>a</sup>	3.0 <sup>b</sup>	3.0 <sup>b</sup>
	Organic matter (% soil dry weight)	2.2 <sub>2,112</sub>	0.1119	14.9	13.5	17.5
	Carbon <sup>*</sup> (% soil dry weight)	2.1 <sub>2,112</sub>	0.0131	8.3 <sup>a</sup>	7.9 <sup>a</sup>	10.0 <sup>b</sup>
	Nitrogen (% soil dry weight)	2.9 <sub>2,112</sub>	0.0617	0.3	0.2	0.3
	Soil C to N ratio <sup>***</sup> (%)	8.4 <sub>2,112</sub>	<0.001	29.7 <sup>b</sup>	34.1 <sup>a</sup>	32.3 <sup>a</sup>
Fruiting bodies	Dry weight (g)	3.1 <sub>2,68</sub>	0.0513	1.0	0.3	0.6
	Abundance <sup>**</sup>	5.0 <sub>2,68</sub>	0.0097	1.9 <sup>b</sup>	2.0 <sup>ab</sup>	2.7 <sup>a</sup>
	Shannon's Diversity Index <sup>***</sup>	10.8 <sub>1,113</sub>	<0.001	0.4 <sup>b</sup>	0.8 <sup>a</sup>	0.8 <sup>a</sup>
	Species richness <sup>***</sup>	12.0 <sub>2,113</sub>	<0.001	2.0 <sup>b</sup>	3.2 <sup>a</sup>	3.3 <sup>a</sup>
Soil fungi	Shannon's	7.8 <sub>2,115</sub>	<0.001	5.0 <sup>a</sup>	4.9 <sup>b</sup>	4.8 <sup>b</sup>
	Species richness <sup>***</sup>	8.2 <sub>1,115</sub>	<0.001	137.7 <sup>a</sup>	130.5 <sup>b</sup>	126.6 <sup>b</sup>

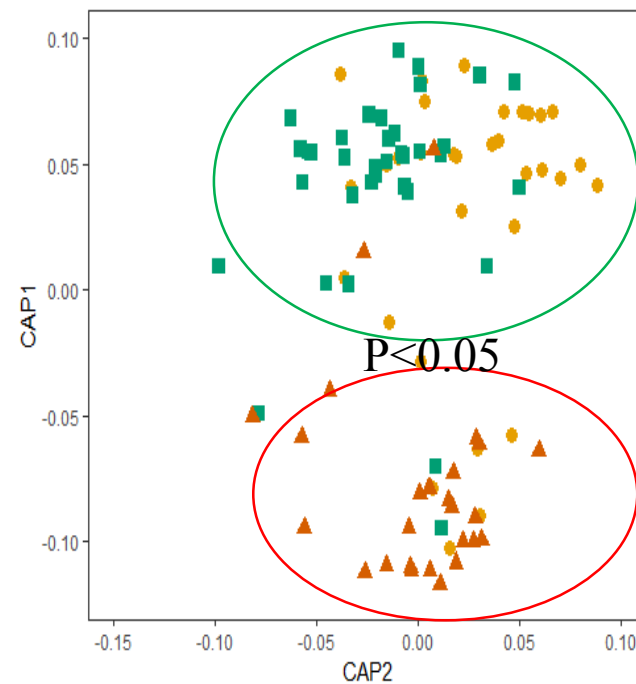
# PERMANOVA Models

- Continuous cover
- Clear-fell
- Unmanaged

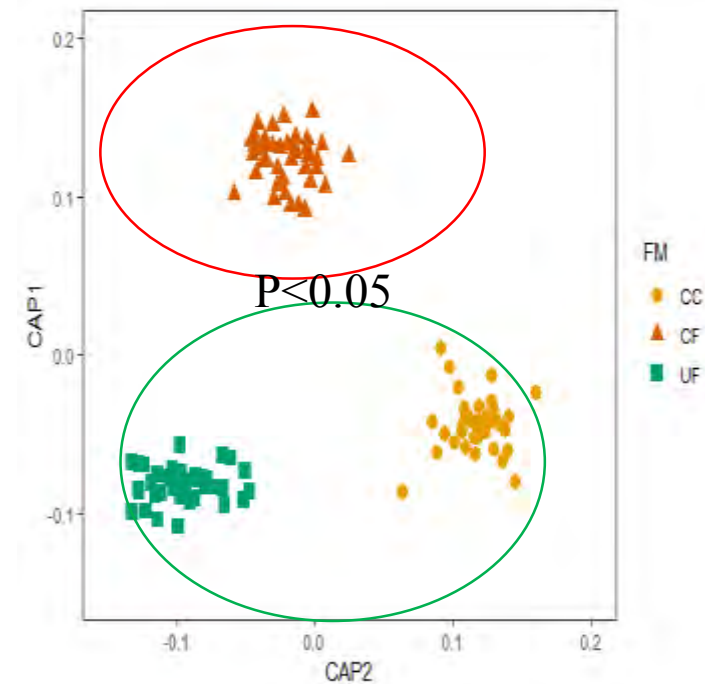
2) The main effect and pair-wise PERMANOVA models – > Canonical Analysis of Principal Coordinates



Fruiting Body Dry Weight



Fruiting Body Abundance



Soil Fungi

# Similarity Percentage (SIMPER) Analysis

Species	Group CCF Av.Abund d	Group CF Av.Abund d	Av.Diss	Diss/SD	Contrib %	Cum.%
<i>Microphale perforance</i>	35.51	1.41	17.75	1.05	18.04	18.04
<i>Cortinarius sp.</i>	1.69	7.06	4.16	0.37	4.23	22.27
<i>Mycena sp.</i>	3.61	3.40	3.34	0.35	3.39	25.66
<i>Laccarius fulvissimus</i>	0.00	5.97	2.98	0.28	3.03	28.69
<i>Marasmius alliaceus</i>	5.62	0.00	2.81	0.33	2.86	31.54
<i>Mycena galopus</i>	4.98	0.74	2.77	0.37	2.82	34.36
<i>Hygrophoropsis aurantiaca</i>	0.00	5.28	2.64	0.37	2.68	37.04
<i>Collybia peronata</i>	0.95	4.24	2.53	0.28	2.57	39.62
<i>Mycena fagetorum</i>	2.93	1.63	2.23	0.24	2.26	41.88
<i>Mycena epipterygia</i>	3.33	1.28	2.22	0.35	2.26	44.14
<i>Thelephora terrestris</i>	0.00	4.41	2.21	0.28	2.24	46.38
<i>Trichaptum abietinum</i>	1.95	2.10	1.97	0.23	2.00	48.39
<i>Gloeophyllum sepiarium</i>	0.00	3.92	1.96	0.27	1.99	50.38

Fruiting body Abundance

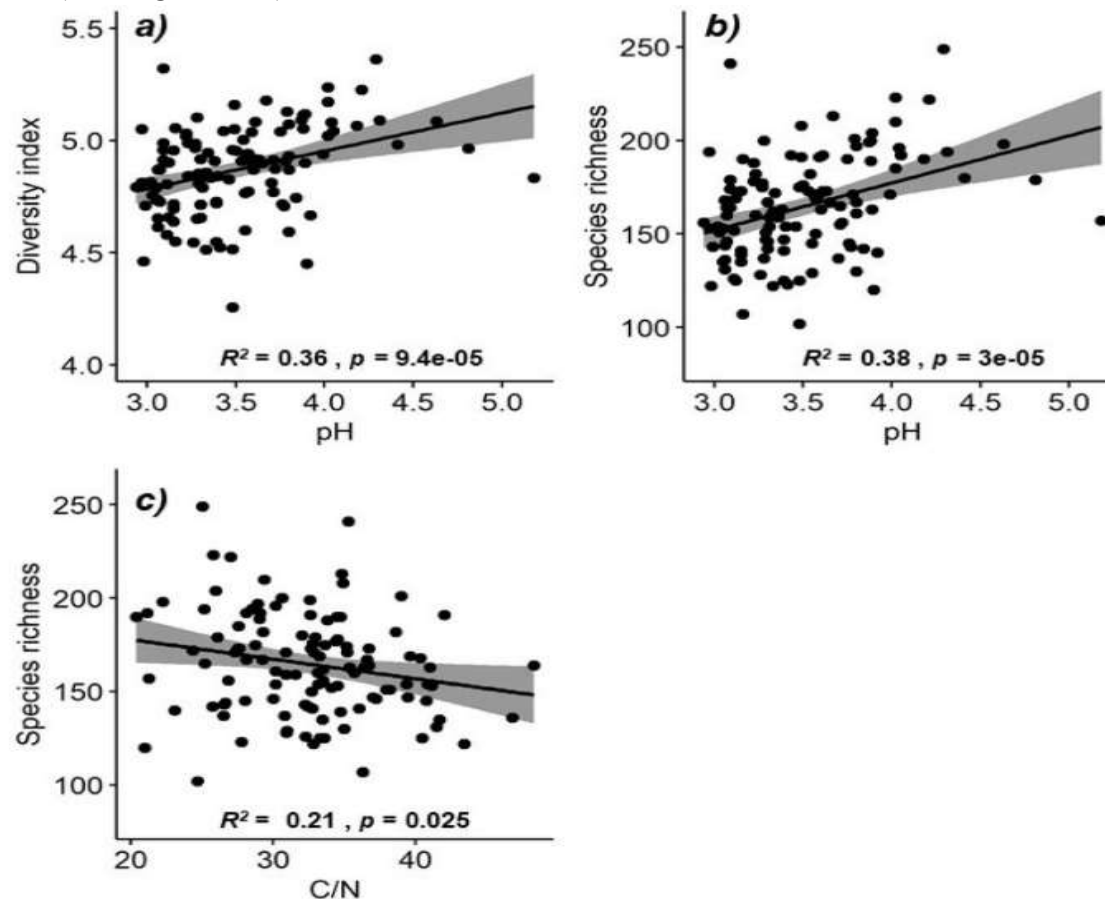
Species	Rank	Group CC Av.Abund	Group CF Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
<i>Hyaloscyphaceae</i>	Family	0,87	1,03	0,53	1,14	0,72	0,72
<i>Thelephora terrestris</i>	Species	0,1	1,05	0,53	0,81	0,71	1,43
<i>Luellia</i>	Genus	0,61	0,92	0,5	1,07	0,67	2,1
<i>Venturiaceae</i>	Family	0,87	0,74	0,49	1,12	0,65	2,75
<i>Archaeorhizomyces</i>	Class	0,27	0,82	0,48	0,51	0,64	3,39
<i>Piloderma sphaerosporum</i>	Species	0,98	0,22	0,46	1,05	0,62	4,01
<i>Archaeorhizomyces</i>	Class	0,92	0,41	0,45	1,04	0,61	4,62
<i>Unknown Fungi (34)</i>	Kingdom	0,87	0,3	0,43	0,86	0,58	5,2
<i>Archaeorhizomyces</i>	Class	1,1	0,49	0,43	1,02	0,57	6,34
<i>Penicillium simile</i>	Species	0,47	1,2	0,4	1,59	0,54	7,43
<i>Piloderma</i>	Genus	0,8	0,06	0,4	0,93	0,53	7,96
<i>Solicoccozyma terricola</i>	Species	0,95	1,21	0,39	1,28	0,53	8,49
<i>Umbelopsis</i>	Genus	0,8	0,86	0,39	1,3	0,52	9,02
<i>Penicillium_austroafricanum</i>	Species	1,1	1,45	0,39	1,28	0,52	9,54
<i>Hyaloscyphaceae</i>	Family	1,21	0,7	0,38	1,22	0,52	10,05
<i>Geomyces asperulatus</i>	Species	1,17	1,02	0,38	1,19	0,51	10,56
<i>Cryptococcus neoformans</i>	Species	0,54	0,52	0,37	1	0,49	11,05
<i>Oidiodendron pilicola</i>	Species	1,14	1,04	0,36	1,2	0,48	11,54
<i>Meliniomyces variabilis</i>	Species	0,98	0,71	0,35	1,4	0,47	12,01
<i>Ascomycota</i>	Phylum	0,81	0,77	0,35	1,26	0,47	12,48
<i>Apiotrichum sporotrichoides</i>	Species	0,68	0,92	0,34	0,93	0,46	12,94
<i>Cortinarius stillatitius</i>	Species	0,64	0,04	0,33	0,41	0,44	13,38
<i>Leucosporidiales</i>	Order	0,53	0,71	0,32	1,23	0,43	13,81
<i>Hyaloscyphaceae</i>	Family	0,53	0,39	0,32	0,81	0,42	14,66
<i>Mortierella macrocystis</i>	Species	1,14	0,9	0,28	1,39	0,38	15,04

Soil Fungi

# Correlation Analysis

## 3) Variation of soil fungal communities by soil chemical properties

Adjusted P-value=0.001. *Italic*: significant ( $P < 0.05$ ) before Bonferroni correction.  
ns(non-significant) > 0.003, \* < 0.003



## Correlation with soil chemical properties

*Soil fungal diversity*

Strong relationship with soil pH.

Close to significant: C to N ratio

*Fruiting body diversity*

None.

# Distance-based Linear Models

## 3) Variation of soil macro-micro fungal communities by soil chemical properties

Dataset	Variable	Pseudo-F	P-Value	Prop.
Fruiting body abundance	pH**	2.0	<b>0.002</b>	3.0%
	OM*	1.5	<b>0.033</b>	2.2%
	C	1.4	0.11	2.0%
	N	1.3	0.142	1.9%
	C/N**	1.9	<b>0.009</b>	2.9%
Fruiting body dry weight	pH*	1.4	<b>0.036</b>	2.1%
	OM	1.1	0.273	1.6%
	C	1.0	0.4	1.6%
	N	1.1	0.312	1.6%
	C/N*	1.4	<b>0.027</b>	2.2%
Soil fungi	pH***	7.3	<b>0.001</b>	6.1%
	OM***	2.0	<b>0.001</b>	1.8%
	N**	1.8	<b>0.002</b>	1.5%
	C***	2.1	<b>0.001</b>	1.8%
	C/N***	5.3	<b>0.001</b>	4.5%

> **Fungal communities – Soil chemistry**

Fruiting body – pH& C/N +(OM)

Soil fungi– All soil variable

P-value: ns > 0.05, \* < 0.05, \*\* < 0.01, \*\*\* < 0.001.

# Main Findings

⇒ **Continuous-cover forestry maintains similar soil chemical properties and fungal communities to Unmanaged forest.**

⇒ Forest Management Practices significantly altered soil chemical properties, fruiting body and soil fungal diversity.

⇒ CCF retained similar soil pH and C/N to UF than CF.

## Why CF is different?



1. Moisture and sun influx
2. Environmental disturbances (soil erosion, ECM fungi)

Dataset	Variable	Clear fell	Continuo us cover	Unmanaged	$F_{(df)}$	P-value
Soil Chemistry	pH***	3.24 <sup>a</sup>	3.03 <sup>b</sup>	2.99 <sup>b</sup>	12.9 <sub>2,114</sub>	<0.001
	OM	14.9	13.5	17.5	2.2 <sub>2,112</sub>	0.1119
	Carbon*	8.27 <sup>a</sup>	7.93 <sup>a</sup>	10.02 <sup>a</sup>	2.1 <sub>2,112</sub>	0.0131
	Nitrogen	0.28	0.23	0.31	2.9 <sub>2,112</sub>	0.0617
	C/N***	29.69 <sup>b</sup>	34.11 <sup>a</sup>	32.29 <sup>a</sup>	8.4 <sub>2,112</sub>	<0.001

# Unexpected Findings



## ➤ Fruiting body diversity



Clear-fell

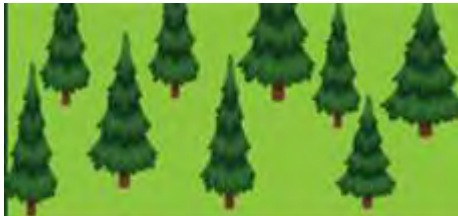


Continuous-cover



Unmanaged

## ➤ Soil fungal diversity



Unmanaged



Continuous-cover



Clear-fell

## Why?!

- Logging residues and stumps
- Soil scarification

- Dormant & external seeds
- Replanted species

# Key Species

- Continuous-cover forestry maintains similar soil fungal communities to Unmanaged forest

Top 3 fruiting bodies: *Micromphale perforans*, *Hygrophoropsis aurantiaca* and *Thelephora terrestris*



Decompose coniferous needle



Appears near fallen trees and tree stumps



Abundant in regenerated stands

Top 3 soil fungi: *Piloderma sphaerosporum*, *Thelephora terrestris* and *Luellia sp*



Dominant in late decaying stage



Abundant in regenerated stands



Associated with *Orthilia secunda*

# Relationships

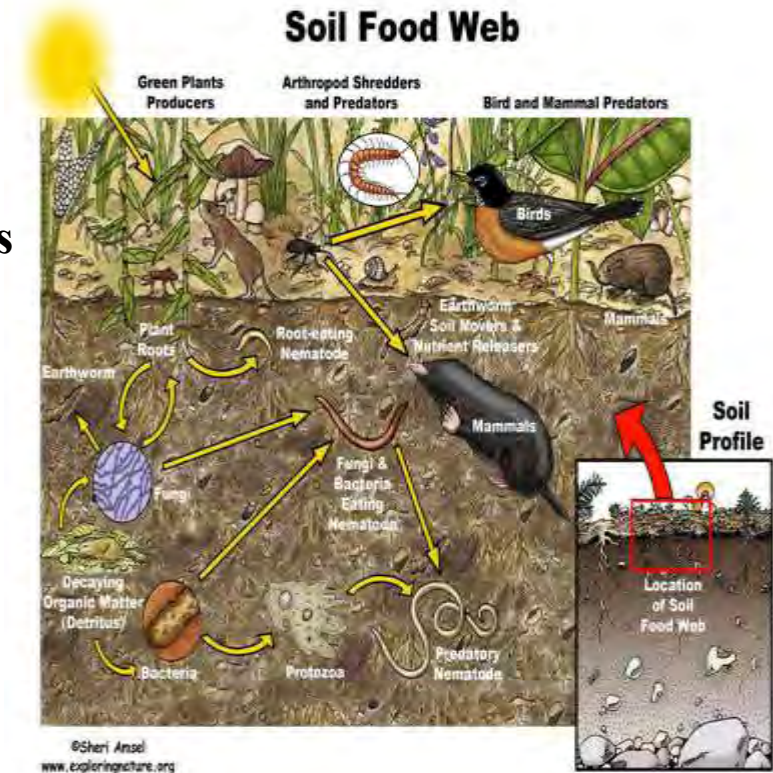
➤ Variation in soil fungal communities is related to soil chemical properties

-Fruiting body --- Soil pH, C/N (4.3~5.9%)+ abiotic factors

-Soil fungi ---- Soil pH, C/N, C, N (15.7%) + abiotic factors

-Correlation with Fruiting body: mushroom is a fraction of fungal communities

-Correlation with Soil fungi: soil pH --- diversity \*\*\* soil C/N --- diversity \*



## Take Home Message

- **Mimicking natural disturbance during harvest may reduce impacts on forest ecosystems.**
- **Clear-felling had pronounced effect on biological and chemical properties of soils.**
- **Partial harvest of 30% maintained similar fungal communities to unmanaged forests.**
- **Variation in soil fungal communities correlated to soil chemical properties.**
- **Continuous-cover forestry is a promising option for maintaining soil ecosystems.**



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## Forest Ecology and Management

Volume 480, 15 January 2021, 118659



# Continuous-cover forestry maintains soil fungal communities in Norway spruce dominated boreal forests

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# Acknowledgement

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- Delphine Noel
- Georgina Mary
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